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THE STRUCTURE OF ARCHANGELOLIDE, A SESQUITERPENIC LACTONE FROM Laserpitium archangelica Wulf.

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From the roots of Laserpitium archangelica WULF. β -sitosterol, laserin (II), 3,4-methylenedioxy-5methoxypropiophenone (III), and a new sesquiterpenic lactone — archangelolide — were isolated. The latter was also isolated from the fruits of the same species. On the basis of the PMR spectra of the native substance and their derivatives, as well as on the basis of chemical correlation with acetylisomontanolide (IV), structure I was inferred for archangelolide.

From the roots of several species of *Laserpitium* a series of sesquiterpenic substances has been isolated¹⁻¹¹. In this paper we describe substances from the roots and the rhizomes of *L. archangelica* WULF. (*Umbelliferae.*) From the dry residue of the light petroleum extract substance *I* crystallised out, the IR and PMR spectra of which indicated that it is an as yet undescribed sesquiterpenic γ -lactone to which we gave the name archangelolide. On chromatography of the crude extract we also isolated in addition to archangelolide (*I*) the already described laserin⁴ (*II*), 3,4-methylenedioxy-5-methoxypropiophenone^{2,12-15} (*III*),** and β -sitosterol, which were identified by comparison with authentic samples.

Archangelolide (I), of m.p. $69-71^{\circ}$ C and $[\alpha]_{D} + 35\cdot8^{\circ}$, displayed in its IR spectrum absorption bands of a γ -lactone group (1791 and 1150 cm⁻¹), a saturated ester group (1738 cm⁻¹), and α , β -unsaturated ester group (1713 cm⁻¹), a double bond (1651 cm⁻¹), and an acetate group (1240 cm⁻¹). Its PMR spectrum (Table I) indicated the presence of an O-angelyl group, an O- α -methylbutyryl group, and two acetyl groups. Its mass spectrum indicated the presence of a fragment of maximum mass m/e 488, further fragments of m/e 388 (488–100), 326 (488–102–60), 286 (488–100–102), and 226 (488–100–102–60). In accordance with the PMR and IR spectra. CO⁺), 55 (C₄H₇⁺), 57 (C₄H₉⁺), 83 (C₄H₇·CO⁺), 85 (C₄H₉·CO⁺) and 100 (C₄H₇·COOH).

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^{**} For this compound we use chemical nomenclature and consider both trivial names, crocatone^{14,15} and latifolone^{4,9} as superfluous.



In view of the presence of the fragments of m/e 226 and 326, indicating the elimination of a single molecule of acetic acid, it was possible to assume that the second O-acetyl group, which was demonstrated by the PMR spectrum, could be easily eliminated, and that the fragment of m/e 488 is pseudomolecular M' = M - 60, similarly as in acetylisomontanolide¹⁰ (IV), for example. The mass M = 548 corresponds to the elemental composition $C_{29}H_{40}O_{10}$ for archangelolide (I). By hydrogenation of archangelolide (1) on PtO_2 in acetic acid we obtained compound V which according to its elemental analysis, PMR, IR and mass spectrum was a saturated tetraester of the composition $C_{29}H_{44}O_{10}$, containing two O-acetyl groups and two α -methylbutyryl groups in the molecule. From these facts it followed that the molecule of archangelolide (I) contained two acetate groups, one residue of angelic acid, one of a-methylbutyric acid, and a basic fifteencarbon bicyclic skeleton with one double bond and a γ -lactone ring. The character of the skeleton was deduced directly from a detailed analysis of the PMR spectrum of archangelolide (I) by multiple resonance experiments (Table I). From the observed distribution of the chemical shifts and the topological continuity of vicinal and long-range couplings (Table I) of protons it followed that its skeleton is a 3-guaiene with three esterified secondary hydroxyls, on $C_{(2)}$, $C_{(6)}$, and $C_{(8)}$, and with two esterified tertiary hydroxyls, on C(10) and C(11). The observed values of vicinal couplings of protons on the fragment $C_{(5)}$ to $C_{(9)}$ (Table I) indicated the same relative configuration (i.e. 5α -H, 6β -H, 7α -H, 8β -H) and hence also the same conformation

of the seven-membered rings as in montanolide⁹ (VI), isomontanolide¹⁰ (VII), and acetylisomontanolide¹⁰ (IV); hence, the orientation of the γ -lactone ring at C₍₆₎ is also the same.



These conclusions were confirmed by direct chemical correlation of archangelolide (I) with acetylisomontanolide¹⁰ (IV). Hydrogenation of archangelolide (I) in acetic acid on 30% palladized charcoal gave tetrahydroarchangelolide (V) and two triester lactones, VIII and IX, of the same composition $C_{24}H_{34}O_8$, which according to their PMR spectra (Table I) contained two O-acetyl groups and one a-methylbutyryl group, bound to $C_{(10)}$, $C_{(11)}$, and $C_{(8)}$. The saturated triester lactone IX was identical according to its IR, PMR, and mass spectrum with the tetrahydroacetylisomontanolide¹⁰ (IX) described earlier. Triester lactones VIII and IX differed only by the configuration of the methyl group at $C_{(4)}$. Relative configurations of this group were assigned on the basis of the determined magnitudes of average vicinal coupling of methyl protons $H_{(15)}(\overline{J}_{4,15} = 7.2 (VIII) \text{ and } \overline{J}_{4,15} = 6.8 (IX))$, from which the 4 β configuration of the methyl in triester VIII and the 4 α configuration of this group in compound IX follows on the basis of the rule for ${}^{3}J_{(CH_{1}-CH)}$ on the CH₃--CH-CH' fragment ${}^{3}J_{CH_{3}}$ (H,H' anti-periplanar) > ${}^{3}J_{CH_{3}}$ (H,H' syn-periplanar), under the supposition of α -configuration of the proton H₍₅₎ (ref.^{16,17}). From the formation of tetrahydroacetylisomontanolide¹⁰ (IX) from archangelolide (I) an unambiguous localisation of the acetyl groups in its molecule followed, i.e. on C(10) and $C_{(11)}$, as well as the localisation of one bound C_5 -acid residue esterifying the hydroxyl at $C_{(8)}$. The decision, which of the two C_5 -acid residues is bound to $C_{(2)}$ and which to C(8), was carried out in the following manner: On oxidation of archangelolide (I) with m-chloroperbenzoic acid we obtained a mixture of compounds from which we isolated a small amount of epoxide X and a fraction XI containing according to its PMR and mass spectrum a mixture of substances with the retained Δ^3 double bond and variously oxidized angelic acid residue. We hydrogenated this fraction XI without further separation under conditions suitable for hydrogenolysis, as mentioned in the section on hydrogenolysis of archangelolide (I).

From the mixture of hydrogenation products we isolated a product, $C_{24}H_{34}O_8$, which according to its PMR spectrum was a mixture of triester lactones *VIII* and *IX*.

 H_{7}

			-	-	
I ^b	3.32		3·03 ^c	4.80	3.62
	$J_{1,2} 3.5$			$J_{6,5} 11.5$	$J_{7,6} 9.7$
	J _{1,5} 80			J _{6,7} J ¹	J _{7,8} 11.4
V ^e	3.13	$2 \cdot 30^d$	$2 \cdot 10^d$	4.62	3.61
	$J_{1,2}$ 7.5			$J_{6.5} 11.5$	$J_{7.6} 10$
	J _{1,5} 7.5			$J_{6,7}^{-10.0}$	$J_{7,8} \ 10.5$
VIII ^f	_	2.60^{d}	1.85 ^d	4.60	3.60
				$J_{6,5}$ 12.1	$J_{7,6} 10.2$
				$J_{6,7}$ 10.2	$J_{7,8} \ 11.2$
ryh	· · ·	2.204	2.254	4.00	2.55
IA	_	2.30	2.23	4·90	J- 10.0
				$J_{6,7} = 12.0$	$J_{7,6}$ 100 $J_{7,6}$ 11.2
				0,5	7,0
X ⁱ	3.00		2.64	4.76	3.65
	$J_{1,2}$ 7.9		$J_{5,6}$ 12.2	$J_{6.5}$ 12.2	$J_{7.6}$ 9.9
			7.1	1 9.8	10.9

TABLE I

Compound⁴

H.

Characteristic Parameters of the PMR Spe	ectra of Archangelolide and Its Derivatives
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He

H

H.

From the course of the mentioned hydrogenolysis it followed unambiguously that the residue of angelic acid is at $C_{(2)}$, and that of α -methylbutyric acid, at $C_{(8)}$. The sum of the mentioned facts shows that the structure with a partial relative stereostructure represented by formula *I* should be assigned to archangelolide. The sterical arrangement on the chirality centers at $C_{(1)}$, $C_{(5)}$, $C_{(6)}$, $C_{(7)}$, $C_{(8)}$, $C_{(10)}$, and $C_{(11)}$ in archangelolide (*I*) is the same as in montanolide⁹ (*VI*), isomontanolide¹⁰ (*VII*), and acetylisomontanolide¹⁰ (*IV*).

The structure of archangelolide (I) represents a further example of a relatively rich substitution by oxygen-containing functions of the basic skeleton of sesquiterpenic lactones present in the species of *Laserpitineae* tribe (*Umbelliferae*); this was stated already in a preceding paper¹⁰. Moreover, archangelolide (I) is the first and only tetraester described in the field of sesquiterpenic lactones so far. In analogy to the majority of sesquiterpenic lactones which were already isolated from the species

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TABLE I

(Continued)

H ₈	H ₉	H _{9'}	H ₁₃	H ₁₄	H ₁₅
5.61 $J_{8,9} = 2.7$ $J_{8,9}, 11.0$ $J_{8,7} = 11.0$	$2.60 \\ J_{9,8} 2.7 \\ J_{9,9}, 15$	2·04 ^d	1.56	1.42	$ \begin{array}{r} 1 \cdot 93 \\ J_{15,3} & \pm \ 0 \\ J_{15,5} & \pm \ 0 \\ J_{15,2} & \pm \ 0 \end{array} $
5.55 $J_{8,9} < 2$ $J_{8,9'}, 10.5$ $J_{8,7}, 10.5$	2·52 J _{9,9} , 15	1·92 ^d J _{9',8} 10·5	1.52	1.52	1·10 J _{15,4} 7·0
$5.53 \\ J_{8,9} \leq 1 \\ J_{8,9}, 10 \\ J_{8,7}, 11$	2·48 9,9', 15	1·85 J _{9',9} 15 J _{9',8} 10	1.51 ^g	1.53 ^g	0·99 J _{15,4} 7·2
$5.47 \\ J_{8,9} 2.5 \\ J_{8,9} 10.9 \\ J_{8,7} 10.9 $	2·49 J _{9,9} , 15	1·96 J _{9',9} 15 J _{9',8} 10·5	1.51	1.51	1·12 J _{15,4} 6·8
$5.44 \\ J_{8,9} 2.6 \\ J_{8,9}, 10.6 \\ J_{8,7} 10.6 $	2.57 $J_{9,8}$ 2.5 $J_{9,9}$, 15	2·02 ^d	1.54 ^{<i>g</i>}	1.54 ^g	1.63 ^{<i>g</i>}

^aAll spectra were measured on a Varian HA-100 instrument using deuteriochloroform as a solvent and tetramethylsilane as an internal reference. Chemical shifts are given in δ [TMS)-scale and splittings in Hz (first-order analysis). The assignments of signals were confirmed by decoupling experiments. ^bO-Angelyl – β -H: 6:04 qq; O- α -methylbutyryl – α -H: 2:29 m ($J \approx 7$), α -CH₃: 1:14 d ($J \approx 7$) and β -CH₃: 0:92 t ($J \approx 7$); O-acetyls – 2:00 and 2:06 (s); H₂ 5:82; H₃ 5:65; ^cBroad doublet of doublets, ³ $J \approx 20$ Hz. ⁴Approximate position from DR-experiments. ⁶Spectrum obscured, due to impurities; O- α -methylbutyryl – α -H and α -H': 2:30 (DR), α -CH₃: 1:12 d ($J \approx 7$), α -CH₃: 1:11 d ($J \approx 7$), β -CH₃: 0:92 t (J?) and β -CH₃: 0:93 t ($J \approx 7$); O-acetyls: 2:04 and 2:10 (s). ^aTentative assignment. ^bO- α -Methylbutyryl: α -CH₃: 1:14 d ($J \approx 7$) and β -CH₃: 0:92 t ($J \approx 7$); O-acetyls: 2:02 and 2:05 (s). ⁱ O- α -Methylbutyryl: α -CH₃: 1:14 d ($J \approx 7$) and β -CH₃: 0:92 t ($J \approx 7$); O-acetyls: 2:06 and 1:99 (s); O-angelyl: β -H: 6:08 qq; H₂ 5:29, $J_{2,.1} = 80$, $J_{2,.3} = 1:8$; H₃ = 3:56, $J_{3,.2} = 1:7$. of the mentioned tribe, in archangelolide too the oxygen-containing substituent is at $C_{(11)}$. The structure of archangelolide (I) is also interesting because its basic sesquiterpenic moiety has the same structure, including the configuration and conformation, as montanolide⁹ (VI), isomontanolide¹⁰ (VII), and acetylisomontanolide¹⁰ (IV), but its angelic acid residue is bound to the hydroxyl at $C_{(2)}$ and not at $C_{(8)}$ as is the case in compounds VI, VII and IV. Considering the localisation of the α -methylbutyryl residue in the position $C_{(8)}$ and the angelyl residue in the α -position to the skeletal double bond, and disregarding other similarities and differences, archangelolide shows a certain structural analogy with trilobolide^{6.7} (XII) which we isolated from Laser trilobum (L.) BORKH., closely related to the species of Laserpitium. The presence of lactone I in L. archangelica is additional evidence of a certain relationship between Umbelliferea and Compositae, because the sesquiterpenic lactones are a typical component of the species of Compositae.

EXPERIMENTAL

Melting points were determined on a Kofier block and they were not corrected. For column chromatography silica gel according to Pirra and Sitrica¹⁴ (30-60 µ, deacuivated by addition of 11% of water) was used. For thin layer chromatography silica gel G Merck was-used. IR spectra were measured in chloroform on a Unicam SP 200 spectrophotometer. PMR spectra were measured in deuteriochloroform, unless stated otherwise, on a Varian HA-100 appratus. Mass spectra were measured on an AEI MS 902 spectrograph. Optical rotation was determined with an objective polarimeter in methanol.

Isolation of Substances

A) Roots: Dry, ground roots (2·4 kg) of Laserpitium archangelica WULF, collected in June 1965 on the southern and south-eastern slopes close to the top of Revañ mountain (Martinské hole, Slovakia), were extracted with light petroleum. From the residue archangelolide (I) crystallised out, m.p. $69-71^{\circ}C$ (ether), $[a]_{2}^{O}+35\cdot8^{\circ}$. For $C_{29}H_{40}O_{10}$ (548-6) calculated: $(53\cdot49^{\circ}_{c}C, 7\cdot35^{\circ}_{c}H]$; found: $63\cdot70^{\circ}_{c}C, 7\cdot62^{\circ}_{c}H$. The residue (35 g) was chromatographed on silica gel (350 g) with benzene, giving a mixture of laserin⁴ (II) and 3,4-methylenedioxy-5-methoxypropiophenone² (III) After distillation of the mixture *in vacuo* (approx. 1 Torr) substance III crystallised out, m.p. $88-89^{\circ}C$ (light petroleum), which melted undepressed on admixture of an authentic sample and also had identical infrared spectrum. The liquid fraction after the separation of III was purified by column chromatography on silica gel, giving liquid laserin⁴ (II) identical according to its IR spectrum with an authentic specimen. From a later fraction β -sitosterol was isolated, m.p. 137°C (methanol), which was identified on the basis of its spectrum and mixture melting point with a standard.

B) Fruits: Dry, ground fruits (1.3 kg), collected in July 1971 in the same locality as above were extracted with light petroleum and concentrated. From the residue archangelolide (1) crystallised out, m.p. $69-71^{\circ}$ C (ether), identical according to its IR spectrum and mixture melting point with the substance isolated under A).

Tetrahydroarchangelolide (V)

A mixture of archangelolide (*I*; 1·42 g), 50 ml acetic acid, and 131 mg of PtO₂ was saturated with hydrogen; consumption was 182 ml of hydrogen (25°C, 738 Torr). Conventional work-up gave tetrahydroarchangelolide (*V*; 1·29 g), m.p. 136–137°C (diisopropyl ether), $[\alpha]_{\rm D}$ – 54·3° (*c* 0·14).

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 R_F 0.35 (light petroleum-acetone-moist ether 5:1:1). For $C_{29}H_{44}O_{10}$ (552·6) calculated: 63·08% C, 8·02% H; found: 63·23% C, 8·09% H. IR spectrum: 2950, 1780 (v-lactone), 1725 (saturated ester), 1465, 1370, 1245 (acetate), 1145, 1100, 1075, 1020 cm⁻¹. Mass spectrum: m/e 492 (552 - 60), 450 (552 - 102), 390 (552 - 102 - 60), 330 (552 - 102 - 60 - 60), 228 (552 - 102 - 60 - 60 - 102), 85 (C_4H_9 , CO⁺), 57 ($C_4H_9^+$) (metastable 38·2), and 60 (CH₃COOH).

Hydrogenolysis of Archangelolide

A mixture of archangelolide (I; 532 mg), 20 ml of acetic acid, 121 mg of 30% Pd/C catalyst, and a few crystals of p-toluenesulfonic acid was saturated with hydrogen for 4 days. After conventional working up a product (450 mg) was obtained which according to thin-layer chromatography contained at least two substances. It was chromatographed on a silica gel column (50 g), using benzene with 5% ether as eluent. Triester VIII was eluted first, m.p. 118-120°C (diisopropyl ether), $R_F 0.4$ (light petroleum-acetone-moist ether 5:2:2). Mass spectrum: m/e 392 (452 - 60), 350 (452 - 102), 290 (452 - 102 - 60), 230 (452 - 102 - 60 - 60), 85 (C_4H_9,CO^+) , 57 $(C_4H_9^+)$ (metastable 38.2), and 60 (CH₃COOH). IR spectrum: 2920, 1774 (y-lactone), 1725 (saturated ester), 1465, 1380, 1245 (acetate), 1145, 1095, 1080, 1020 cm⁻¹. Almost simultaneously with the triester VIII the saturated triester IX was also eluted, m.p. 129–131°C (diisopropyl ether), $[\alpha]_D - 41.8^\circ$ (c 0.25). $R_F 0.4$ (light petroleum-acetone-moist, ether 5 : 2 : 2). Mass spectrum: m/e 350 (452 - 102), 332 (452 - 60 - 60), 290 (452 - 102 - 60). 230 (452 - 102 - 60 - 60), 85 (C_4H_9.CO $^+$), 57 (C_4H_9^+) (metastable 38·2) and 60 (CH_3COOH), IR spectrum: 2950, 1775 (y-lactone), 1725 (saturated ester), 1470, 1370, 1250 (acetate), 1145, 1105, 1075, 1020 cm⁻¹. Mixture melting point of this compound (*IX*) with an authentic specimen of tetrahydroacetylisomontanolide¹⁰ (IX) was undepressed. In further fractions tetrahydroarchangelolide (V) of m.p. 145°C (disopropyl ether), came out $[\alpha]_D - 56.6^{\circ}$ (c 0.5), and $R_F 0.35$ (light petroleum-acetone-moist ether 5 : 2 : 2), identical according to its IR spectrum and mixture melting point with the substance obtained in the preceding experiment.

Epoxidation of Archangelolide (I)

A mixture of archangelolide (*I*; 1·0 g), 250 ml of chloroform, and 335 mg of 85% *m*-chlorobenzoic acid was refluxed for 90 minutes. The solvent was distilled off and the product (1·0 g) chromato-graphed on a column of 100 g of silica gel using benzene with 5% of ether as eluent. First fractions contained in addition to the starting compound also traces of epoxide X. Medium fractions afforded the product XI (480 mg), m.p. $60-63^{\circ}$ C, $[a]_{D}-69\cdot7^{\circ}$ (c 0·42). R_{F} 0·35 (benzene-ether 4 : 1). For C₂₉H₄₀O₁₁ (564·6) calculated: 61·68% C, 7·14% H; found: 61·30% C, 7·37% H. Mass spectrum: m/e 448 (564 – 116), 388 (564 – 116 – 60), 286 (564 – 116 – 60 – 102), 226 (564 – 116 – 60) – 102 – 60), 85 (C₄H₉.CO⁺), 57 (C₄H₉⁺) (metastable 38·2). IR spectrum: 3000, 1780 (γ -lactone), 1725 (saturated ester), 1525, 1445, 1430, 1380, 1370, 1330, 1240 (acetate), 1145, 1100, 1015, 925 cm⁻¹.

Hydrogenolysis of product XI. A mixture of product XI (180 mg), 20 ml of acetic acid, 80 mg of 30% Pd/C catalyst, and a crystal of p-toluenesulfonic acid was saturated with hydrogen for 3 days. After working up the mixture in a conventional manner a product (160 mg) was oblained which was chromatographed on silica gel (15 g) using benzene with 10% of ether for elution. In the first fractions a mixture of tetrahydroacetylisomontanolide VIII and IX, m.p. 114–116°C and R_p 0-6 (benzene-ether 4 : 1) appeared. Mass and IR spectra were identical with those of tetrahydroacetylisomontanolide¹⁰ (IX) and the mixture melting point with an authentic specimen was undepressed. In medium fractions a non-crystalline product was eluted, with R_p 0-3 (benzene-

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ether 1: 1). Mass spectrum: m/e 566, 506 (566 - 60), 446 (566 - 60 - 60), 404 (556 - 60 - 102), 288 (566 - 102 - 116 - 60), 228 (566 - 116 - 60 - 60), 85 (C₄H₉.CO⁺), 57 (C₄H₉⁺) (metastable 38·2), 71 (C₄H₉O⁺). IR spectrum: 3000, 2900, 2820, 1780 (γ -lactone), 1730 (saturated ester), 1525, 1460, 1374, 1335, 1245 (acctate), 1150, 1100, 1045, 1020, 985, 935.

Elemental analyses were carried out in the analytical department of our Institute (head Dr J. Horáček) by Mrs V. Rusová and Mr V. Štérba. The IR spectra were measured by Mrs S. Holubová and optical rotations by Mrs J. Větrovská. The mass spectra were measured by Dr A. Trka, Dr J. Kohoutová, and Dr K. Ubik (head Dr L. Dolejš). To all those mentioned, as well as to Miss H. Němcová (technical assistance) we express our sincere thanks.

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